

# Diagnostic Accuracy of Novel and Traditional Rapid Tests for Influenza Infection Compared With Reverse Transcriptase Polymerase Chain Reaction

## A Systematic Review and Meta-analysis

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**Background:** Rapid and accurate influenza diagnostics can improve patient care.

**Purpose:** To summarize and compare accuracy of traditional rapid influenza diagnostic tests (RIDTs), digital immunoassays (DIAs), and rapid nucleic acid amplification tests (NAATs) in children and adults with suspected influenza.

**Data Sources:** 6 databases from their inception through May 2017.

**Study Selection:** Studies in English, French, or Spanish comparing commercialized rapid tests (that is, providing results in <30 minutes) with reverse transcriptase polymerase chain reaction reference standard for influenza diagnosis.

**Data Extraction:** Data were extracted using a standardized form; quality was assessed using QUADAS-2 (Quality Assessment of Diagnostic Accuracy Studies 2) criteria.

**Data Synthesis:** 162 studies were included (130 of RIDTs, 19 of DIAs, and 13 of NAATs). Pooled sensitivities for detecting influenza A from Bayesian bivariate random-effects models were 54.4% (95% credible interval [CrI], 48.9% to 59.8%) for RIDTs, 80.0% (CrI, 73.4% to 85.6%) for DIAs, and 91.6% (CrI, 84.9% to 95.9%) for NAATs. Those for detecting influenza B were 53.2%

(CrI, 41.7% to 64.4%) for RIDTs, 76.8% (CrI, 65.4% to 85.4%) for DIAs, and 95.4% (CrI, 87.3% to 98.7%) for NAATs. Pooled specificities were uniformly high (>98%). Forty-six influenza A and 24 influenza B studies presented pediatric-specific data; 35 influenza A and 16 influenza B studies presented adult-specific data. Pooled sensitivities were higher in children by 12.1 to 31.8 percentage points, except for influenza A by rapid NAATs (2.7 percentage points). Pooled sensitivities favored industry-sponsored studies by 6.2 to 34.0 percentage points. Incomplete reporting frequently led to unclear risk of bias.

**Limitations:** Underreporting of clinical variables limited exploration of heterogeneity. Few NAAT studies reported adult-specific data, and none evaluated point-of-care testing. Many studies had unclear risk of bias.

**Conclusion:** Novel DIAs and rapid NAATs had markedly higher sensitivities for influenza A and B in both children and adults than did traditional RIDTs, with equally high specificities.

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Influenza viruses cause yearly epidemics of acute respiratory illness affecting 5% to 30% of the population (1, 2). Diagnosing influenza on the basis of only clinical symptoms is difficult because manifestations vary and are nonspecific (3). Consequently, results of diagnostic tests are useful to guide clinical management. Potential benefits of rapid and accurate diagnosis of influenza infection include prompt initiation of antiviral therapy (4–6), fewer ancillary diagnostic tests (7, 8), fewer hospitalizations (4, 9), prompt institution of hospital infection control measures (10), and less unnecessary antibiotic use (7, 11).

Diagnostic tests for influenza identify the virus in a patient's respiratory secretions by isolation in cell culture, detection of viral RNA by nucleic acid amplification, or detection of viral antigens by immunoassay (10). Reverse transcriptase polymerase chain reaction (RT-PCR) has replaced viral culture as the gold standard for influenza diagnosis because of its superior analytic and clinical sensitivity (12, 13). However, specimens for RT-PCR are typically sent to specialized laboratories, and testing is done in batches, resulting in turnaround times that extend beyond the clinical encounter. Rapid

influenza diagnostic tests (RIDTs) that detect viral antigens by immunoassay are widely used because they are simple enough to do at the point of care and provide results in less than 30 minutes. A 2012 systematic review and meta-analysis by Chartrand and colleagues evaluated 159 diagnostic accuracy studies of RIDTs published to December 2011 (14). They showed that commercially available RIDTs had high specificity (98.2% [95% CI, 97.5% to 98.7%]) but poor sensitivity (62.3% [CI, 57.9% to 66.6%]). In light of these findings, regulators and professional societies have questioned the utility of RIDTs (13, 15–17). Since 2011, the following 2 novel classes of rapid influenza diagnostic assays (that is, with results available in <30 minutes) have been

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commercialized, with claims of improved sensitivities based on technological improvements: automated immunochromatographic antigen detection tests (digital immunoassays [DIAs]) and rapid nucleic acid amplification tests (NAATs). Digital immunoassays use an instrument-based digital scan of the test strip to enhance antigen detection accuracy by eliminating the need for an operator to visualize and subjectively interpret test results (16). Rapid NAATs use a modified RT-PCR (18) or isothermal amplification technology (19) to greatly reduce analytic times.

In February 2017, the U.S. Food and Drug Administration (FDA) reclassified and instituted minimum performance standards for rapid influenza tests (17). Therefore, an updated and comprehensive synthesis of the evidence on their accuracy is warranted. The primary objective of this systematic review and meta-analysis was to estimate and compare the diagnostic accuracy of commercialized RIDTs, DIAs, and rapid NAATs for detecting influenza A and B infection in patients with suspected influenza, compared with an RT-PCR reference standard. We also aimed to evaluate patient, test, and methodological factors associated with test accuracy within each of the 3 classes of rapid tests.

## METHODS

We used methods recommended by the Cochrane Diagnostic Test Accuracy Working Group (20, 21), including the preparation of a prespecified protocol and analysis plan developed according to the PRISMA-P (Preferred Reporting Items for Systematic reviews and Meta-analyses Protocols) statement (22). The PRISMA guidelines were used for preparing this report (23).

### Data Sources and Searches

On the basis of the PubMed search strategy in Chartrand and colleagues' systematic review (14) and in collaboration with a medical librarian (G.G.), we searched PubMed, Embase, BIOSIS Previews, Scopus, Web of Science, and the Cochrane Central Register of Controlled Trials on 18 August 2015, with an update on 21 May 2017 (Supplement Table 1, available at [Annals.org](#)). We used EndNote (Clarivate Analytics) libraries from Chartrand and colleagues' systematic review (14) to exclude records they had screened and excluded while keeping studies they had included, provided the reference standard used was RT-PCR. In addition, we hand-searched recent guidelines, narrative reviews, and citations of included articles.

### Study Selection

We included peer-reviewed studies in English, French, or Spanish providing original data on the diagnostic accuracy of rapid influenza tests against an RT-PCR reference standard. Eligible participants were children and adults with clinically suspected influenza during periods of influenza activity. Editorials, letters to the editor, and conference abstracts were excluded

because they contain insufficient information on important data items for investigating sources of heterogeneity and ascertaining methodological quality. Studies using a case-control design (spectrum bias) and those performing the reference standard depending on index test results (partial verification bias) were also excluded. We attempted to contact authors if studies provided insufficient information to construct a 2 × 2 table.

Rapid influenza tests were defined as commercially developed assays that detect influenza A, B, or A/B within 30 minutes by identifying influenza viral antigen or RNA directly from an unprocessed specimen. Acceptable specimens included nasopharyngeal aspirates, swabs, or washes; nasal aspirates, swabs, or washes; and throat swabs. For a study to be eligible, the index test and comparator needed to test the same clinical specimen or 2 specimens taken concurrently from the same anatomical site. Commercial and laboratory-developed RT-PCR assays were acceptable reference tests. When more than 1 RT-PCR assay was used as a reference standard, preference was given to the commercial assay with the best reported analytic sensitivity for influenza A. We excluded studies if the rapid test itself was part of a composite reference standard (incorporation bias).

Two reviewers (R.W. and J.M.) independently screened citations (titles and abstracts) identified by our search strategy and not already screened by Chartrand and colleagues. Potentially relevant articles were retrieved in full and screened for eligibility by the 2 reviewers. Disagreements were resolved by consensus or by involvement of a third reviewer (J.P.).

### Data Extraction and Quality Assessment

A data extraction sheet (Supplement Table 2, available at [Annals.org](#)) based on the form used by Chartrand and colleagues was created in Google Forms to minimize the risk for transcriptional errors (24). It was then pilot-tested on a subset of included articles by 2 reviewers (J.M. and R.W.) before being finalized. Two reviewers independently extracted data. Disagreements were resolved by consensus or by a third reviewer (J.P.). Articles that assessed several index tests against a reference standard were counted as several studies; a separate extraction form was completed for each index test.

The study population was considered pediatric or adult if 85% of persons were below or above, respectively, an age cutoff of 18 to 21 years (as defined by the study). In studies that provided separate results for children and adults, we used the age cutoff applied by the investigators. Point-of-care testing was defined as index testing done outside the traditional laboratory setting by persons other than trained laboratory personnel. We considered a study to have been industry-sponsored if a commercial entity funded it or provided index tests.

Two reviewers independently assessed the quality of individual studies using QUADAS-2 (Quality Assessment of Diagnostic Accuracy Studies 2) criteria (25).

## Data Synthesis and Analysis

For each study, we calculated sensitivity, specificity, and positive and negative likelihood ratios (LRs) along with 95% CIs. Results are presented separately for influenza A and B and for each of the 3 index test types (RIDTs, DIAs, and rapid NAATs). We considered influenza A and B to be separate diagnostic targets; studies that reported only combined influenza A/B data were not included in the quantitative synthesis. We calculated the pooled accuracy estimates (sensitivity, specificity, and LR) across studies with 95% credible intervals (CrIs) using Bayesian bivariate random-effects meta-analysis models (details in the **Supplement**, available at [Annals.org](http://Annals.org)). The bivariate random-effects approach deals with potential sources of variation caused by imprecision of sensitivity and specificity estimates within individual studies, correlation between sensitivity and specificity across studies, and variation in sensitivity and specificity between studies (26). Because heterogeneity is expected in meta-analysis of diagnostic accuracy studies, a random-effects model is preferred (21). Analyses were done using noninformative priors. We also used the model to create a plot depicting the pooled estimates with the credible and prediction regions and a hierarchical summary receiver-operating characteristic (HSROC) curve (20, 27, 28).

We first assessed heterogeneity by visual inspection of the HSROC curves and the credible and prediction regions (20). Subgroup analyses were planned to further investigate heterogeneity for covariates that provided at least 3 studies per stratum by index test type. Variables selected a priori as potential sources of heterogeneity were population age (children vs. adults), duration of symptoms before testing, type of respiratory specimen, point-of-care testing, commercial brand, infecting virus subtype, study quality, and industry sponsorship. Summary sensitivity and specificity estimates were calculated for each level of a covariate, along with their 95% CrIs. Differences in accuracy were then compared across levels of a covariate. Analyses were done using STATA, version 13 (StataCorp); R, version 3.2.1 (R Foundation; [www.r-project.org](http://www.r-project.org)); and WinBUGS, version 1.4.3 (29).

Researchers usually assume that RT-PCR is a perfect reference standard (that is, 100% sensitivity and 100% specificity) when doing a meta-analysis of the diagnostic accuracy of comparator tests for respiratory viruses (14, 30, 31). However, acknowledging that accuracy varies across commercial and laboratory-developed RT-PCR assays for influenza (32, 33), we did a sensitivity analysis to assess whether our study conclusions would remain unchanged if we allowed RT-PCR to be considered imperfect. We thus repeated our pooled accuracy calculations without forcing a sensitivity and specificity of 100% for the reference standard in the random-effects models (34, 35).

## Role of the Funding Source

This study was supported in part by the Québec Health Research Fund and by an investigator-initiated

study grant from BD Diagnostic Systems. Funding sources had no involvement in study design, conduct, analysis, or publication.

## RESULTS

### Search Results

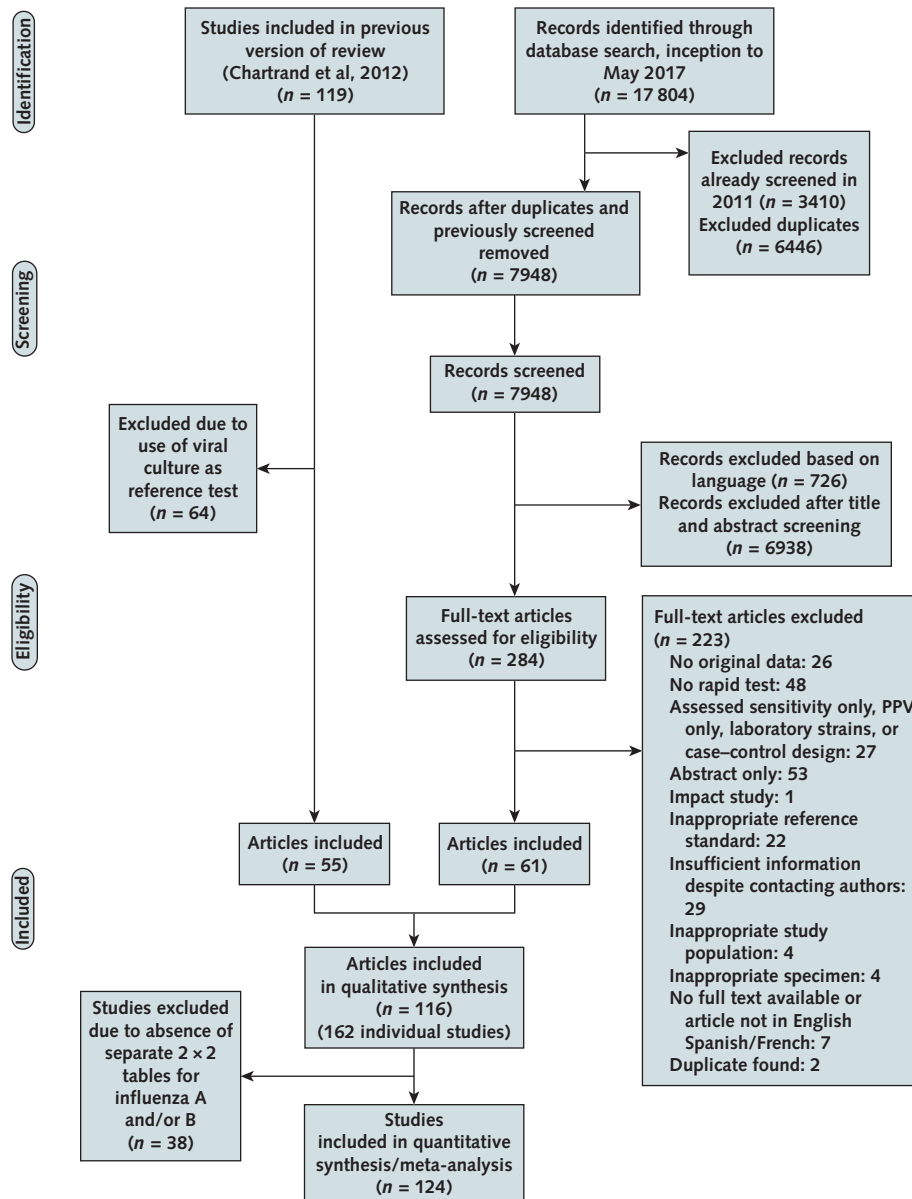
After screening titles and abstracts and doing full-text assessments (**Figure 1**), we included 61 articles (18, 19, 36–67–68–94). Another 55 articles from Chartrand and colleagues' review (14) that used RT-PCR as the reference standard (95–126–127–149) were also included. Of the 162 studies, 38 did not report 2 × 2 tables separately for influenza A and B. Thus, we included 124 studies in our meta-analysis of accuracy estimates.

### Study Characteristics

**Table 1** describes the 162 included studies. **Supplement Table 3** (available at [Annals.org](http://Annals.org)) provides more details of their characteristics and accuracy estimates. Of the studies, 130 (80.2%) investigated the accuracy of traditional RIDTs, 19 (11.7%) of DIAs, and 13 (8.0%) of rapid NAATs. Among RIDTs, 35 commercial tests were evaluated (**Supplement Table 4**, available at [Annals.org](http://Annals.org)). We also evaluated 2 DIAs, the BD Veritor System for Flu A+B (6 studies; 31.6%) (BD Diagnostic Systems) and the Sofia Influenza A+B Fluorescent Immunoassay (13 studies; 68.4%) (Quidel), and 2 rapid NAATs, the Alere i Influenza A & B (8 studies; 61.5%) (Alere) and the cobas Liat Influenza A/B assay (5 studies; 38.5%) (Roche Diagnostics). Most studies assessed mixed populations of adults and children. The population was considered pediatric in 22.3% (29 of 130), 31.6% (6 of 19), and 7.7% (1 of 13) of RIDT, DIA, and rapid NAAT studies, respectively. Point-of-care testing was done in 23.1% of RIDT studies versus 36.8% of DIA and 0% of rapid NAAT studies. Most studies combined patients in hospital and outpatient settings without separating data or did not report the study setting. Nasopharyngeal swabs were the most commonly used specimens (range, 28.5% to 53.8%). Industry sponsorship was more frequent in DIA (68.4%) and rapid NAAT (61.5%) studies than RIDT studies (20.0%). An insufficient number of studies reported on the duration of symptoms before presentation and testing (2 of 15 DIA studies [68, 75] and 1 of 11 NAAT studies [18]).

### Quality Assessment

Quality assessments using QUADAS-2 criteria are summarized in **Supplement Figure 1** (available at [Annals.org](http://Annals.org)). Most RIDT (53.8%) and rapid NAAT (69.2%) studies did not present clear patient or specimen selection criteria and processes or were at high risk of bias; risk of selection bias was less common in DIA studies (42.1%). Limited reporting of blinding to reference standard results during interpretation of the index test resulted in a risk of bias in 15.8% to 63.2% of studies across index test types. Because DIAs and rapid NAATs have machine-based, objective readers, a

**Figure 1.** Study search and selection flow chart.

Flow chart summarizing evidence search and study selection. PPV = positive predictive value.

lack of blinding when evaluating these test results represents a smaller risk of bias than for nonautomated colorimetric assays, such as RIDTs.

## Synthesis of Results

### Primary Analysis: Overall Accuracy

All index test types showed large variability in sensitivity for both influenza A and B across studies (see forest plots in Figures 2 and 3 and HSROC plots in Supplement Figure 2 [available at Annals.org]), whereas specificity was consistently above 95% (Supplement Figure 3, available at Annals.org). Pooled sensitivities and specificities for influenza A and B are presented in Table 2 (see Supplement Table 5, available at Annals.org, for 95% prediction intervals). Forest plots of

individual and pooled LRs are presented in Supplement Figure 4 (available at Annals.org). Because pooled specificity was at least 98.3% across classes, we deemed that any differences between groups would not be clinically relevant. Therefore, we calculated differences in pooled accuracy only for sensitivity. Digital immunoassays had sensitivities that were 25.5 percentage points (95% CrI, 17.0 to 33.4 percentage points) and 23.5 percentage points (CrI, 7.7 to 37.9 percentage points) higher than those for traditional RIDTs for diagnosing influenza A and B, respectively. Rapid NAAT sensitivity was superior to that of RIDTs by 37.1 percentage points (CrI, 28.6 to 44.2 percentage points) and 41.7 percentage points (CrI, 28.5 to 54.0

percentage points) for influenza A and B, respectively, and to that of DIAs by 11.5 percentage points (CrI, 2.9 to 19.5 percentage points) and 18.2 percentage points (CrI, 6.9 to 30.6 percentage points).

### Subgroup Analyses and Sensitivity Analysis: Investigation of Heterogeneity

We did subgroup analyses to explain the heterogeneity in test accuracy (in terms of sensitivity) seen on visual inspection of the forest and HSROC plots (Table 2). Pooled rapid test sensitivity was consistently higher in children than adults (range of differences in sensitivity, 12.1% [CrI, 3.1% to 22.1%] to 31.8% [CrI, 6.1% to 52.6%]), except with rapid NAATs for influenza A

(difference, 2.7% [CrI, -10.7% to 19.7%]). For NAATs, adult-specific pooled estimates were generated from 4 studies, 3 of which were on the Alere assay. We therefore did a post hoc sensitivity analysis removing the Liat study (76). The sensitivities for Alere among adults were 80.3% (CrI, 63.7% to 90.8%) and 68.5% (CrI, 40.2% to 87.2%) for influenza A and B, respectively (Supplement Table 6, available at [Annals.org](http://Annals.org)).

Doing the index test at the point of care did not affect test accuracy for RIDTs and DIAs. No studies evaluated rapid NAATs at the point of care. Pooled sensitivity estimates favored industry-sponsored studies by 6.2 to 34.0 percentage points. Studies evaluating the BD Veritor showed higher pooled sensitivities than

**Table 1.** Characteristics of the 162 Included Studies

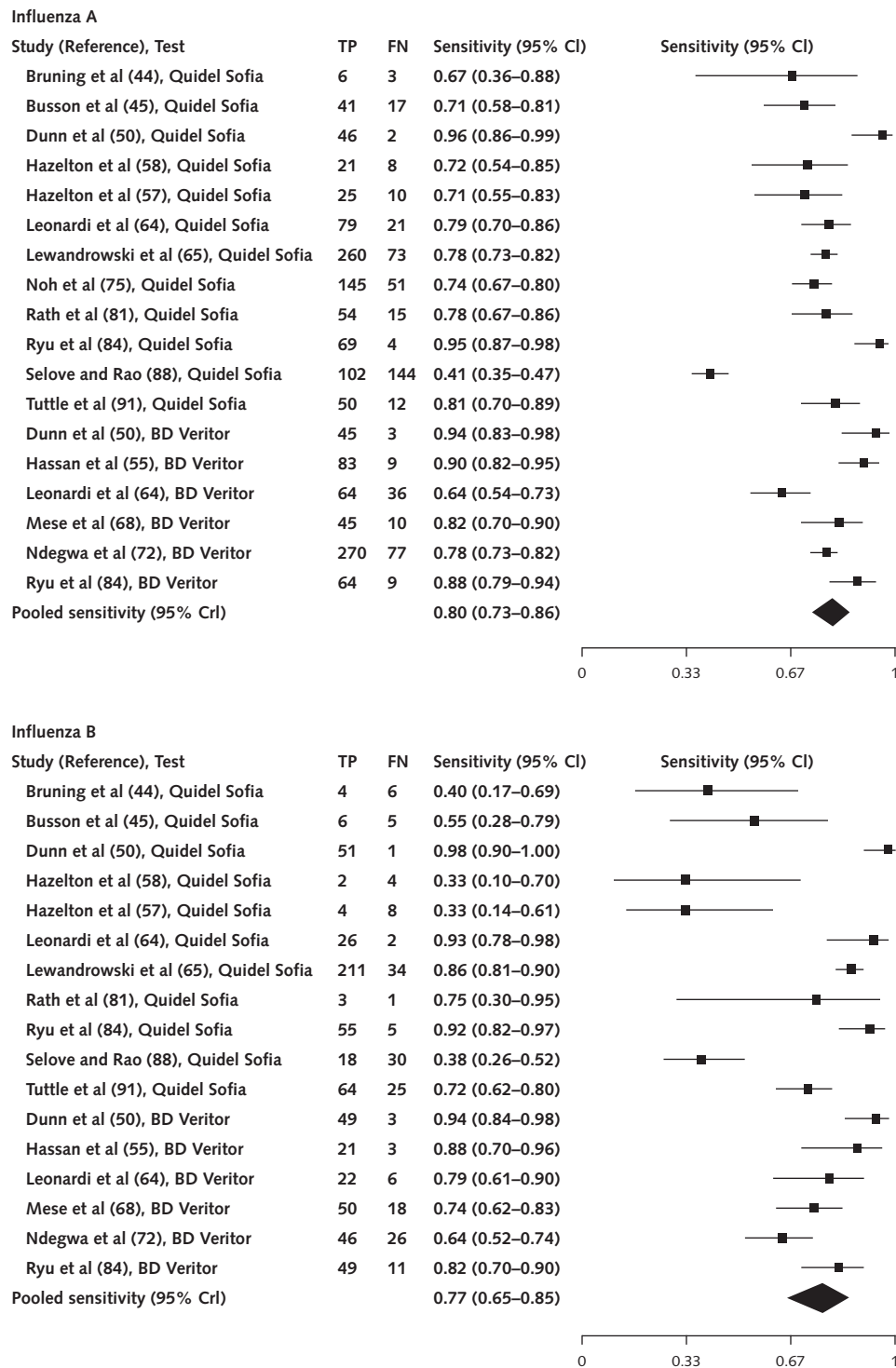
Study Characteristic	Studies of Traditional RIDTs (n = 130), n (%)	Studies of DIAs (n = 19), n (%)	Studies of Rapid NAATs (n = 13), n (%)
<b>Population</b>			
Children	29 (22.3)	6 (31.6)	1 (7.7)
Adults	17 (13.1)	4 (21.1)	1 (7.7)
Mixed/not reported	84 (64.6)	9 (47.4)	11 (84.6)
<b>Commercial brand*</b>			
Directigen Flu A+B	29 (22.3)	-	-
BinaxNOW Influenza A&B	21 (16.2)	-	-
QuickVue Influenza A+B	21 (16.2)	-	-
QuickVue Influenza Test	9 (6.9)	-	-
BD Veritor Flu A+B	-	6 (31.6)	-
Sofia Influenza A+B	-	13 (68.4)	-
Alere i Influenza A & B	-	-	8 (61.5)
Cobas Liat Influenza A/B	-	-	5 (38.5)
<b>Industry sponsorship</b>			
Yes	26 (20.0)	13 (68.4)	8 (61.5)
<b>Majority specimen type</b>			
Nasopharyngeal aspirate or wash	20 (15.4)	0 (0)	0 (0)
Nasopharyngeal swab	37 (28.5)	10 (52.6)	7 (53.8)
Nasal aspirate or wash	7 (5.4)	3 (15.8)	0 (0)
Nasal swab	19 (14.6)	1 (5.3)	1 (7.7)
Throat swab	4 (3.1)	0 (0)	0 (0)
Mixed	7 (5.4)	2 (10.5)	3 (23.1)
Not reported	36 (27.7)	3 (15.8)	2 (15.4)
<b>Setting in which the test was performed</b>			
Outpatient only	31 (23.8)	1 (5.3)	1 (7.7)
Hospital only	15 (11.5)	1 (5.3)	0 (0)
Emergency department only	7 (5.4)	2 (10.5)	0 (0)
Mixed	32 (24.6)	6 (31.6)	6 (46.2)
Other†	3 (2.3)	0 (0)	0 (0)
Not reported	43 (33.1)	9 (47.4)	6 (46.2)
<b>Point-of-care testing</b>			
Yes	30 (23.1)	7 (36.8)	0 (0)
<b>Studies conducted during 2009 H1N1 pandemic period</b>			
Yes	77 (59.2)	0 (0)	2 (15.4)
Not reported	1 (0.8)	1 (5.3)	0 (0)
<b>Influenza strains detected</b>			
Influenza A/B combined	54 (41.5)	5 (26.3)	4 (30.8)
Influenza A	94 (72.3)	18 (94.7)	12 (92.3)
Influenza B	30 (23.1)	17 (89.5)	12 (92.3)

DIA = digital immunoassay; NAAT = nucleic acid amplification test; RIDT = rapid influenza diagnostic test.

\* Among traditional RIDTs, 35 commercially available tests were used (see Supplement Table 4 for list). The 4 most common brands are presented. Note: Directigen Flu A+B includes both Directigen EZ Flu A+B and Directigen Flu A+B.

† Includes nursing homes, Haj pilgrims, and H1N1 school outbreak.

**Figure 2.** Forest plots of the sensitivities of digital immunoassays for influenza A and B.



CrI = credible interval; FN = false negative; TP = true positive.

those evaluating the Quidel Sofia for influenza A (83.0% vs. 77.8%) and B (80.0% vs. 73.5%); however, the 95% CrIs of both differences crossed the null. Test sensitivity was higher for the Liat than the Alere assay for influenza A (97.1% vs. 84.4%; difference, 12.4 percentage points

[CrI, 4.9 to 21.9 percentage points]) and influenza B (98.7% vs. 86.6%; difference, 11.8 percentage points [CrI, 2.8 to 29.5 percentage points]). An insufficient number of studies provided data about the duration of symptoms, type of respiratory specimens used, circulat-

ing subtype, or clinical setting (hospitalized vs. outpatient) to allow for these subgroup analyses.

We did a sensitivity analysis for our overall pooled estimates that relaxed the assumption that RT-PCR is a perfect reference standard (Supplement Table 6). As expected, we observed higher accuracy estimates than in our primary analysis for all index test types. Nevertheless, our main finding that DIAs and rapid NAATs demonstrated markedly higher sensitivities for influenza A and B than did traditional RIDTs did not change.

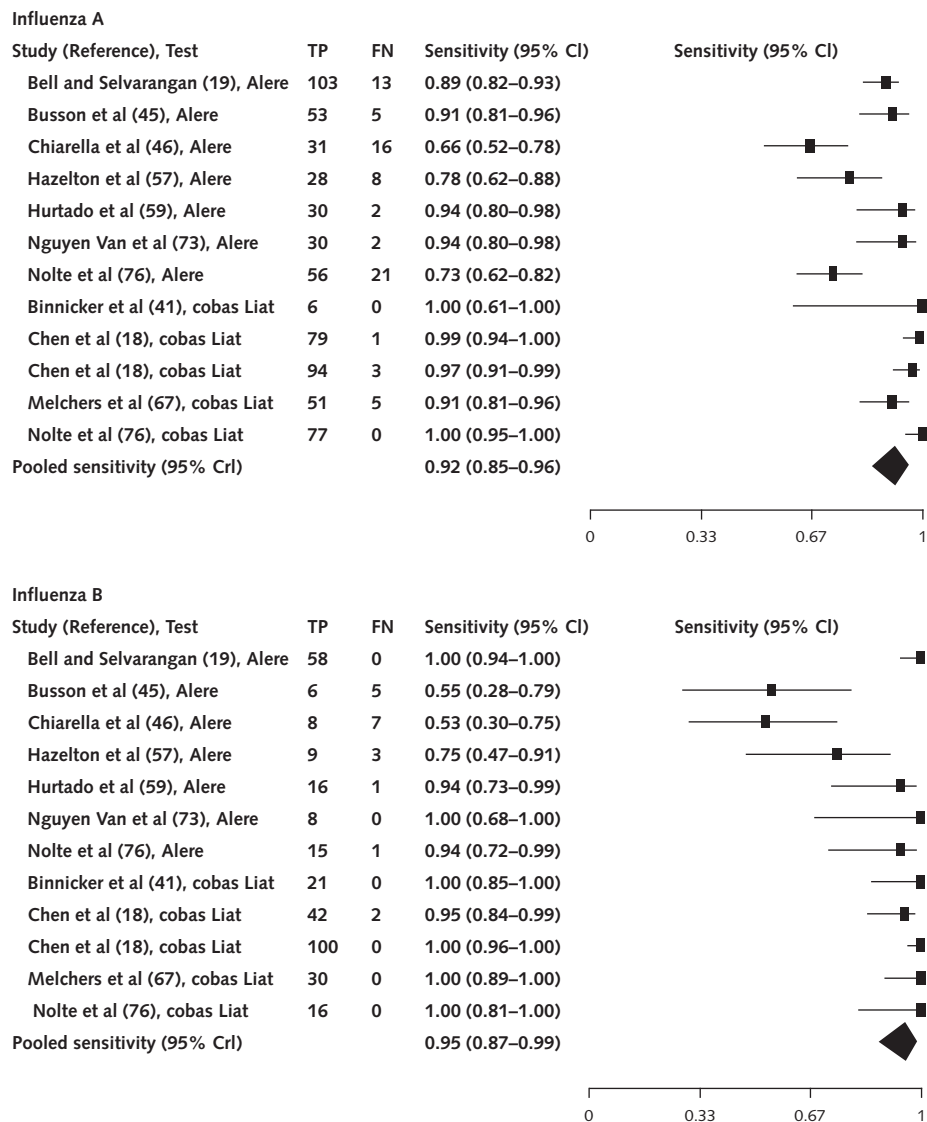
**DISCUSSION**

Through an update and expansion of Chartrand and colleagues' 2012 systematic review and meta-analysis on traditional RIDTs, this study synthesizes the available evidence and compares the diagnostic accu-

acy of commercially available rapid tests for the detection of influenza A and B infection. Like RIDTs, the newer DIAs and rapid NAATs are simple, fast, and approved for use at the point of care by nonlaboratory personnel. Overall, the rapid tests displayed very high specificities ( $\geq 98.3\%$ ) and positive LR ( $>48$ ). Physicians can therefore diagnose influenza with confidence on the basis of a positive RIDT, DIA, or rapid NAAT result. This should lead to improved patient outcomes and decreased health care costs through prompt implementation of infection control measures and initiation of antiviral treatment when indicated, while decreasing unnecessary ancillary investigations and antibiotic over-use (5, 7, 9).

A key finding of our study is that the pooled sensitivities for DIAs (80.0% for influenza A and 76.8% for

**Figure 3.** Forest plots of the sensitivities of rapid nucleic acid amplification tests for influenza A and B.



CrI = credible interval; FN = false negative; TP = true positive.

**Table 2.** Overall and Subgroup Analyses of Pooled Rapid Test Accuracy Estimates for Influenza A and B, by Index Test Type\*

Index Test Type	Influenza A		Influenza B	
	Pooled Sensitivity (95% CrI), %	Pooled Specificity (95% CrI), %	Pooled Sensitivity (95% CrI), %	Pooled Specificity (95% CrI), %
<b>Overall</b>				
Traditional RIDTs (94 influenza A studies; 30 influenza B studies)	54.4 (48.9 to 59.8)	99.4 (99.1 to 99.7)	53.2 (41.7 to 64.4)	99.8 (99.7 to 99.9)
DIA (18 influenza A studies; 17 influenza B studies)	80.0 (73.4 to 85.6)	98.3 (97.4 to 98.9)	76.8 (65.4 to 85.4)	98.7 (97.5 to 99.4)
Rapid NAATs (12 influenza A studies; 12 influenza B studies)	91.6 (84.9 to 95.9)	99.2 (98.6 to 99.7)	95.4 (87.3 to 98.7)	99.4 (98.9 to 99.8)
Difference in sensitivities, overall				
Traditional RIDTs vs. DIAs	<b>-25.5 (-33.4 to -17.0)</b>	-	<b>-23.5 (-37.9 to -7.7)</b>	-
Traditional RIDTs vs. rapid NAATs	<b>-37.1 (-44.2 to -28.6)</b>	-	<b>-41.7 (-54.0 to -28.5)</b>	-
DIAs vs. rapid NAATs	<b>-11.5 (-19.5 to -2.9)</b>	-	<b>-18.2 (-30.6 to -6.9)</b>	-
<b>Subgroup analyses†</b>				
Study population (age)‡				
Traditional RIDTs				
Children (31 influenza A studies; 9 influenza B studies)	61.2 (55.0 to 67.2)	99.2 (98.5 to 99.7)	65.7 (45.3 to 80.5)	99.6 (99.2 to 99.8)
Adults (23 influenza A studies; 5 influenza B studies)	42.6 (34.8 to 50.9)	99.5 (98.6 to 99.8)	33.2 (19.9 to 50.7)	99.9 (99.4 to 100)
Difference in RIDT sensitivity: children vs. adults	<b>18.5 (8.4 to 28.3)</b>	-	<b>31.8 (6.1 to 52.6)</b>	-
DIAs				
Children (11 influenza A studies; 11 influenza B studies)	87.6 (81.8 to 92.2)	98.1 (96.4 to 99.1)	82.5 (71.2 to 90.2)	98.8 (95.6 to 99.7)
Adults (8 influenza A studies; 7 influenza B studies)	75.4 (66.6 to 82.6)	96.7 (94.7 to 98.0)	57.0 (39.5 to 71.6)	98.8 (97.5 to 99.5)
Difference in DIA sensitivity: children vs. adults	<b>12.1 (3.1 to 22.1)</b>	-	<b>25.3 (6.9 to 44.7)</b>	-
Rapid NAATs				
Children (4 influenza A studies; 4 influenza B studies)	90.2 (79.2 to 95.8)	99.0 (96.8 to 99.8)	95.9 (82.9 to 99.2)	99.5 (98.2 to 99.9)
Adults (4 influenza A studies; 4 influenza B studies)	87.4 (71.1 to 95.6)	98.0 (93.2 to 99.5)	75.7 (51.8 to 90.7)	99.3 (97.8 to 99.8)
Difference in NAAT sensitivity: children vs. adults	2.7 (-10.7 to 19.7)	-	<b>19.5 (1.0 to 43.7)</b>	-
Location of testing				
Traditional RIDTs				
POC (17 influenza A studies; 4 influenza B studies)	61.2 (48.4 to 72.7)	98.0 (96.2 to 99.0)	44.8 (17.5 to 76.2)	99.5 (98.9 to 99.8)
Not POC (77 influenza A studies; 26 influenza B studies)	52.9 (46.9 to 58.8)	99.6 (99.3 to 99.8)	54.4 (43.1 to 65.7)	99.9 (99.8 to 99.9)
Difference in RIDT sensitivity: POC vs. not POC	8.2 (-5.9 to 21.3)	-	-9.5 (-39.1 to 23.5)	-
DIAs				
POC (7 influenza A studies; 6 influenza B studies)	77.6 (70.4 to 83.4)	98.1 (95.7 to 99.1)	72.0 (57.4 to 82.0)	98.7 (95.9 to 99.5)
Not POC (11 influenza A studies; 11 influenza B studies)	82.3 (72.1 to 89.7)	98.2 (97.1 to 98.9)	80.4 (64.5 to 90.3)	98.4 (96.5 to 99.4)
Difference in DIA sensitivity: POC vs. not POC	-4.7 (-14.9 to 7.0)	-	-8.4 (-25.9 to 10.0)	-
Industry sponsorship				
Traditional RIDTs				
Sponsored (20 influenza A studies; 8 influenza B studies)	70.8 (60.0 to 79.6)	99.1 (98.2 to 99.6)	77.8 (60.0 to 90.1)	99.7 (99.4 to 99.9)
Not sponsored (74 influenza A studies; 22 influenza B studies)	50.0 (43.9 to 55.2)	99.5 (99.1 to 99.7)	43.5 (33.3 to 54.3)	99.8 (99.7 to 99.9)
Difference in RIDT sensitivity: sponsored vs. not sponsored	<b>21.1 (9.1 to 31.7)</b>	-	<b>34.0 (13.7 to 50.5)</b>	-
DIAs				
Sponsored (13 influenza A studies; 12 influenza B studies)	84.1 (78.3 to 88.8)	98.1 (96.7 to 98.9)	79.7 (66.1 to 88.4)	98.6 (96.0 to 99.5)
Not sponsored (5 influenza A studies; 5 influenza B studies)	67.5 (52.9 to 79.4)	98.6 (97.0 to 99.4)	69.4 (51.1 to 84.2)	99.0 (97.9 to 99.5)
Difference in DIA sensitivity: sponsored vs. not sponsored	<b>16.5 (3.5 to 31.9)</b>	-	10.0 (-9.6 to 30.4)	-

Continued on following page



Table 2—Continued

Index Test Type	Influenza A		Influenza B	
	Pooled Sensitivity (95% CrI), %	Pooled Specificity (95% CrI), %	Pooled Sensitivity (95% CrI), %	Pooled Specificity (95% CrI), %
Rapid NAATs				
Sponsored (8 influenza A studies; 8 influenza B studies)	92.6 (84.5 to 96.8)	99.4 (98.6 to 99.8)	97.2 (92.7 to 99.0)	99.6 (99.1 to 99.8)
Not sponsored (4 influenza A studies; 4 influenza B studies)	86.2 (71.3 to 94.7)	98.5 (96.2 to 99.5)	80.0 (53.0 to 94.7)	98.6 (96.3 to 99.5)
Difference in NAAT sensitivity: sponsored vs. not sponsored	6.2 (−5.1 to 21.7)	−	<b>16.9 (1.7 to 44.1)</b>	−
Commercial brand DIAs				
Sofia (12 influenza A studies; 11 influenza B studies)	77.8 (68.8 to 85.4)	98.5 (97.4 to 99.2)	73.5 (55.8 to 86.1)	98.0 (95.4 to 99.1)
Veritor (6 influenza A studies; 6 influenza B studies)	83.0 (73.4 to 90.1)	97.5 (95.5 to 98.7)	80.0 (68.8 to 88.2)	99.5 (98.8 to 99.8)
Difference in DIA sensitivity: Sofia vs. Veritor	−5.1 (−16.4 to 6.9)	−	−6.4 (−25.8 to 10.4)	−
Rapid NAATs				
Alere (7 influenza A studies; 7 influenza B studies)	84.4 (75.3 to 90.9)	98.9 (97.7 to 99.6)	86.6 (69.0 to 95.3)	99.1 (98.1 to 99.7)
Liat (5 influenza A studies; 5 influenza B studies)	97.1 (92.9 to 98.9)	99.4 (98.4 to 99.8)	98.7 (95.6 to 99.7)	99.5 (98.7 to 99.9)
Difference in NAAT sensitivity: Alere vs. Liat	<b>−12.4 (−21.9 to −4.9)</b>	−	<b>−11.8 (−29.5 to −2.8)</b>	−

CrI = credible interval; DIA = digital immunoassay; NAAT = nucleic acid amplification test; POC = point of care; RIDT = rapid influenza diagnostic test.

\* Differences in pooled sensitivity estimates between groups that did not include the null (0%) in its 95% CrI are in boldface.

† For subgroups that contained  $\geq 3$  studies per stratum by index test type.

‡ Data from studies performed in  $\geq 85\%$  adult or  $\geq 85\%$  pediatric populations or from studies of mixed-age populations that provided data for the adult and pediatric subgroups.

influenza B) and rapid NAATs (91.6% for influenza A and 95.4% for influenza B) are markedly higher than those for RIDTs. The use of DIAs and rapid NAATs improves detection of true cases of influenza by 25 and 40 percentage points, respectively, compared with RIDTs.

Similar to Chartrand and colleagues, we found that traditional RIDTs have summary sensitivities (54.4% for influenza A and 53.2% for influenza B) well below new FDA minimum performance requirements. Effective December 2018, tests detecting influenza antigens will need to show a sensitivity of at least 80%, with a 95% CI lower bound of 70%, against an RT-PCR comparator (17).

Our literature searches to May 2017 identified 1 meta-analysis of rapid influenza test accuracy based on a literature search done in the past 5 years (150). Bruning and colleagues reviewed the accuracy of rapid tests for respiratory viruses against RT-PCR. They included 134 influenza studies (122 RIDTs, 9 Sofia, 1 BD Veritor, and 2 Alere) published to January 2016 and reported pooled sensitivity and specificity estimates of 61.1% and 98.9% for any influenza. They did not evaluate DIAs as a class but saw a pooled sensitivity for the Sofia of 75.3%. Of importance, the literature on novel tests has evolved rapidly since January 2016. Our review included an additional 8 DIA and 10 NAAT studies. We could thus compare results across classes of tests and perform clinically relevant subgroup analyses within each class. Moreover, we presented all data separately for influenza A and B, in keeping with FDA guidance.

The improved sensitivity of DIAs is likely due to proprietary chemistry innovations and to automated readers that eliminate the subjectivity of an operator visualizing and interpreting test results (16). Molecular techniques, such as NAATs, are expected to have low analytic detection limits and thus higher clinical sensitivity (151). We found that rapid NAATs were the only class of rapid tests with overall negative LRs below 0.1, thereby making a negative result useful to rule out influenza (152). However, the cost of DIAs (\$15 to \$20 per test) is similar to that of RIDTs, whereas rapid NAATs may cost 2 to 5 times that amount. Whether the incremental gains in sensitivity of rapid NAATs versus DIAs are worth their added costs will likely depend on the patient populations and clinical contexts in which they are used. Moreover, different commercial rapid NAATs might not perform equally. Pooled sensitivities were above 97% for the Liat modified RT-PCR assay. In contrast, the Alere isothermal assay had sensitivities for influenza A and B of 84.4% and 86.6%, respectively, similar to those of the Veritor DIA (83.0% and 80.0%).

By updating Chartrand and colleagues' review on RIDTs, we could make direct statistical comparisons of the performance of RIDTs versus newer rapid tests. However, we note that no new information has been gained from studying traditional RIDTs since 2012. Despite the addition of 39 evaluations of RIDTs published after Chartrand and colleagues' review, summary estimates for this class are nearly identical. Thus, additional diagnostic accuracy research on RIDTs seems to be of

no value; it will not change future pooled estimates or the interpretation of the test's clinical utility (153).

In children, the pooled sensitivities for influenza A and B of DIAs (87.6% and 82.5%) and rapid NAATs (90.2% and 95.9%) make these newer-generation tests acceptable for use in the pediatric population. We saw that RIDT and DIA sensitivity was higher by approximately 15 percentage points for influenza A and 30 percentage points for influenza B in children than adults, likely related to more prolonged and abundant viral shedding in the former (154, 155). Studies of DIAs in adult populations exhibited pooled sensitivities of at most 75% and negative LR of 0.25 or more. Therefore, clinicians should be aware of the possibility of false-negative results in adults tested by DIA and consider retesting by RT-PCR if the result could influence patient management. In contrast, rapid NAAT pooled sensitivities of 87.4% and 75.7% for influenza A and B in adults suggest that they may be the preferred rapid test in this population. However, we caution that our adult rapid NAAT results should not be overinterpreted because they are based on only 4 studies (442 participants; 155 influenza infections). Moreover, 3 of the 4 adult studies evaluated the Alere assay.

The DIA and rapid NAAT assays evaluated in our review have received Clinical Laboratory Improvement Amendments waivers as low-complexity tests that can be done outside the laboratory. On the basis of 7 studies, we found that using DIAs at the point of care did not affect performance. Unfortunately, no studies evaluating NAATs at the point of care were identified. Given their high sensitivity, NAATs may be prone to false-positive results if protocol breaches cause environmental contamination. Moreover, the Alere NAAT is approved for point-of-care testing only if done directly on a swab. Some included studies evaluated the Alere on swabs eluted in transport medium, for which it is considered a moderately complex assay. Further investigation of the performance, feasibility, and effect of near-patient testing with DIAs and NAATs is therefore warranted; this is the setting in which they are expected to most improve patient outcomes (156, 157).

Studies that declared industry sponsorship produced higher sensitivity estimates by 6.2 to 34.0 percentage points than nonsponsored studies. These differences were statistically significant for RIDTs, DIAs for influenza A, and NAATs for influenza B. This is a consideration when interpreting our results, because a large proportion of DIA (68.4%) and rapid NAAT (61.5%) studies received industry sponsorship in the form of funding or in-kind provision of study materials. This underscores the importance of conducting and publishing independent diagnostic accuracy evaluations of commercial assays, preferably done within the flow of the usual diagnostic pathway, either in the clinical laboratory or at the point of care.

This review has potential limitations. First, we could not assess publication bias because no reliable methods exist to investigate this in diagnostic accuracy studies (20). Second, we saw differences in the distribution of covariates that could affect test sensitivity, such as

point-of-care testing and industry sponsorship, across the 3 index test types. Unfortunately, because of the limited number of studies on DIAs and rapid NAATs, we could not adjust simultaneously for all covariates using metaregression. Moreover, no NAAT studies evaluated point-of-care testing and very few reported adult-specific data. Third, our pooled estimates do not account for the conditional dependence of several index tests done on a single sample. This may lead to underestimation of between-study variance and narrower CIs but would not affect point estimates. Fourth, this study was partly funded by BD Diagnostic Systems, the manufacturers of a DIA. However, we used publicly available data and transparent methods to draw our inferences, and sponsors were not involved in study design, conduct, analysis, or interpretation. Finally, we also highlight a lack of important contextual information in the evidence base. Data not readily available in the diagnostic laboratory, such as study setting, clinical manifestations, presence of comorbid conditions, and duration of symptoms, could affect test accuracy but are frequently missing from reports.

Because of their simplicity and speed, rapid influenza tests are potentially valuable diagnostic tools, especially if deployed at the point of care. Understanding the performance characteristics of different test methods across different patient populations is important for laboratory directors who must decide on their implementation and clinicians who must interpret their results for patient management. The results of our systematic review and meta-analysis, using RT-PCR as a reference standard, suggest that traditional RIDTs are likely to be phased out by regulatory agencies like the FDA because of their poor sensitivity, especially in adults. Digital immunoassays and rapid NAATs showed markedly higher sensitivities for influenza A and B than did RIDTs, with equally high specificities. Performance of the newer-generation rapid influenza tests was also better in pediatric than adult populations, although the difference was less pronounced for rapid NAATs. Additional clinical impact and cost-effectiveness analyses of DIAs and NAATs should help guide decisions about applying rapid testing for influenza in clinical practice.

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**Reproducible Research Statement:** *Study protocol:* See Supplement. *Statistical code:* See Supplement. *Data set:* Full data set and codebook available on [www.nandinidendukuri.com](http://www.nandinidendukuri.com).

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## References

1. Monto AS, Koopman JS, Longini IM Jr. Tecumseh study of illness. XIII. Influenza infection and disease, 1976-1981. *Am J Epidemiol*. 1985;121:811-22. [PMID: 4014174]
2. Monto AS. Epidemiology of viral respiratory infections. *Am J Med*. 2002;112 Suppl 6A:4S-12S. [PMID: 11955454]
3. Nicholson KG, Wood JM, Zambon M. Influenza. *Lancet*. 2003;362:1733-45. [PMID: 14643124]
4. Benito-Fernández J, Vázquez-Ronco MA, Morteruel-Aizkuren E, Mintegui-Raso S, Sánchez-Etxaniz J, Fernández-Landaluce A. Impact of rapid viral testing for influenza A and B viruses on management of febrile infants without signs of focal infection. *Pediatr Infect Dis J*. 2006;25:1153-7. [PMID: 17133161]
5. D'Heilly SJ, Janoff EN, Nichol P, Nichol KL. Rapid diagnosis of influenza infection in older adults: influence on clinical care in a routine clinical setting. *J Clin Virol*. 2008;42:124-8. [PMID: 18289930] doi:10.1016/j.jcv.2007.12.014
6. Blaschke AJ, Shapiro DJ, Pavia AT, Byington CL, Ampofo K, Stockmann C, et al. A national study of the impact of rapid influenza testing on clinical care in the emergency department. *J Pediatric Infect Dis Soc*. 2014;3:112-8. [PMID: 24872879] doi:10.1093/jpids/pit071
7. Esposito S, Marchisio P, Morelli P, Crovari P, Principi N. Effect of a rapid influenza diagnosis. *Arch Dis Child*. 2003;88:525-6. [PMID: 12765923]
8. Iyer SB, Gerber MA, Pomerantz WJ, Mortensen JE, Ruddy RM. Effect of point-of-care influenza testing on management of febrile children. *Acad Emerg Med*. 2006;13:1259-68. [PMID: 17079787]
9. Bonner AB, Monroe KW, Talley LI, Klasner AE, Kimberlin DW. Impact of the rapid diagnosis of influenza on physician decision-making and patient management in the pediatric emergency department: results of a randomized, prospective, controlled trial. *Pediatrics*. 2003;112:363-7. [PMID: 12897288]
10. Renaud C, Papenburg J. Health care-associated viral respiratory tract infections due to influenza virus, respiratory syncytial virus, and other respiratory viruses. In: Leber AL, ed. *Clinical Microbiology Procedures Handbook*. 4th ed. Washington, DC: Am Soc Microbiol Pr; 2016.
11. Noyola DE, Demmler GJ. Effect of rapid diagnosis on management of influenza A infections. *Pediatr Infect Dis J*. 2000;19:303-7. [PMID: 10783019]
12. Centers for Disease Control and Prevention. Guidance for clinicians on the use of RT-PCR and other molecular assays for diagnosis of influenza virus infection. Accessed at [www.cdc.gov/flu/professionals/diagnosis/molecular-assays.htm](http://www.cdc.gov/flu/professionals/diagnosis/molecular-assays.htm) on 6 April 2017.
13. Harper SA, Bradley JS, Englund JA, File TM, Gravenstein S, Hayden FG, et al; Expert Panel of the Infectious Diseases Society of America. Seasonal influenza in adults and children—diagnosis, treatment, chemoprophylaxis, and institutional outbreak management: clinical practice guidelines of the Infectious Diseases Society of America. *Clin Infect Dis*. 2009;48:1003-32. [PMID: 19281331] doi:10.1086/598513
14. Chartrand C, Leeflang MM, Minion J, Brewer T, Pai M. Accuracy of rapid influenza diagnostic tests: a meta-analysis. *Ann Intern Med*. 2012;156:500-11. [PMID: 22371850] doi:10.7326/0003-4819-156-7-201204030-00403
15. World Health Organization. Influenza (seasonal). Accessed at [www.who.int/mediacentre/factsheets/fs211/en](http://www.who.int/mediacentre/factsheets/fs211/en) on 6 April 2017.
16. Dunn JJ, Ginocchio CC. Can newly developed, rapid immunochromatographic antigen detection tests be reliably used for the laboratory diagnosis of influenza virus infections? *J Clin Microbiol*. 2015;53:1790-6. [PMID: 25274999] doi:10.1128/JCM.02739-14
17. Microbiology Devices; Reclassification of Influenza Virus Antigen Detection Test Systems Intended for Use Directly With Clinical Specimens, 82 Fed. Reg. 3609 (Jan. 12, 2017) (to be codified at 21 C.F.R. pt. 866). Accessed at [www.federalregister.gov/documents/2017/01/12/2017-00199/microbiology-devices-reclassification-of-influenza-virus-antigen-detection-test-systems-intended-for](http://www.federalregister.gov/documents/2017/01/12/2017-00199/microbiology-devices-reclassification-of-influenza-virus-antigen-detection-test-systems-intended-for) on 30 March 2017.
18. Chen L, Tian Y, Chen S, Liesenfeld O. Performance of the cobas® Influenza A/B assay for rapid PCR-based detection of influenza compared to Prodesse ProFlu+ and viral culture. *Eur J Microbiol Immunol (Bp)*. 2015;5:236-45. [PMID: 26716012] doi:10.1556/1886.2015.00046
19. Bell JJ, Selvarangan R. Evaluation of the Alere I influenza A&B nucleic acid amplification test by use of respiratory specimens collected in viral transport medium. *J Clin Microbiol*. 2014;52:3992-5. [PMID: 25210070] doi:10.1128/JCM.01639-14
20. Leeflang MM. Systematic reviews and meta-analyses of diagnostic test accuracy. *Clin Microbiol Infect*. 2014;20:105-13. [PMID: 24274632] doi:10.1111/1469-0691.12474
21. Macaskill P, Gatsonis C, Deeks JJ, Harbord RM, Takwoingi Y. Analysing and presenting results. In: Deeks JJ, Bossuyt PM, Gatsonis C, eds. *Cochrane Handbook for Systematic Reviews of Diagnostic Test Accuracy*. Version 1.0. The Cochrane Collaboration; 2010. Accessed at <http://methods.cochrane.org/sdt/handbook-dta-reviews> on 29 March 2017.
22. Moher D, Shamseer L, Clarke M, Ghersi D, Liberati A, Petticrew M, et al; PRISMA-P Group. Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015 statement. *Syst Rev*. 2015;4:1. [PMID: 25554246] doi:10.1186/2046-4053-4-1
23. Liberati A, Altman DG, Tetzlaff J, Mulrow C, Gøtzsche PC, Ioannidis JP, et al. The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate health care interventions: explanation and elaboration. *Ann Intern Med*. 2009;151:W65-94. [PMID: 19622512]
24. Li T, Vedula SS, Hadar N, Parkin C, Lau J, Dickersin K. Innovations in data collection, management, and archiving for systematic reviews. *Ann Intern Med*. 2015;162:287-94. [PMID: 25686168] doi:10.7326/M14-1603
25. Whiting PF, Rutjes AW, Westwood ME, Mallett S, Deeks JJ, Reitsma JB, et al; QUADAS-2 Group. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. *Ann Intern Med*. 2011;155:529-36. [PMID: 22007046] doi:10.7326/0003-4819-155-8-201110180-00009
26. Steingart KR, Schiller I, Horne DJ, Pai M, Boehme CC, Dendukuri N. Xpert® MTB/RIF assay for pulmonary tuberculosis and rifampicin resistance in adults. *Cochrane Database Syst Rev*. 2014:CD009593. [PMID: 24448973] doi:10.1002/14651858.CD009593.pub3
27. Schiller I, Dendukuri D. HSROC: an R package for Bayesian meta-analysis of diagnostic test accuracy. 2015:1-27. Accessed at <https://cran.r-project.org/web/packages/HSROC/vignettes/Tutorial.pdf> on 29 March 2017.

28. Harbord RM, Deeks JJ, Egger M, Whiting P, Sterne JA. A unification of models for meta-analysis of diagnostic accuracy studies. *Biostatistics*. 2007;8:239-51. [PMID: 16698768]
29. Spiegelhalter DJ, Thomas A, Best NG. WinBUGS Version 1.4.3 User Manual. Cambridge, UK: Medical Research Council; 2007.
30. Chartrand C, Tremblay N, Renaud C, Papenburg J. Diagnostic accuracy of rapid antigen detection tests for respiratory syncytial virus infection: systematic review and meta-analysis. *J Clin Microbiol*. 2015;53:3738-49. [PMID: 26354816] doi:10.1128/JCM.01816-15
31. Nicholson KG, Abrams KR, Batham S, Medina MJ, Warren FC, Barer M, et al. Randomised controlled trial and health economic evaluation of the impact of diagnostic testing for influenza, respiratory syncytial virus and *Streptococcus pneumoniae* infection on the management of acute admissions in the elderly and high-risk 18- to 64-year-olds. *Health Technol Assess*. 2014;18:1-274, vii-viii. [PMID: 24875092] doi:10.3310/hta18360
32. Klungthong C, Chinnawirotpisan P, Hussem K, Phonpakobsin T, Manasatienkij W, Ajariyakhajorn C, et al. The impact of primer and probe-template mismatches on the sensitivity of pandemic influenza A/H1N1/2009 virus detection by real-time RT-PCR. *J Clin Virol*. 2010;48:91-5. [PMID: 20413345] doi:10.1016/j.jcv.2010.03.012
33. Salez N, Vabret A, Lueruz-Ville M, Andreoletti L, Carrat F, Renois F, et al. Evaluation of four commercial multiplex molecular tests for the diagnosis of acute respiratory infections. *PLoS One*. 2015;10:e0130378. [PMID: 26107509] doi:10.1371/journal.pone.0130378
34. Dendukuri N, Schiller I, Joseph L, Pai M. Bayesian meta-analysis of the accuracy of a test for tuberculous pleuritis in the absence of a gold standard reference. *Biometrics*. 2012;68:1285-93. [PMID: 22568612] doi:10.1111/j.1541-0420.2012.01773.x
35. Sinclair A, Xie X, Teltscher M, Dendukuri N. Systematic review and meta-analysis of a urine-based pneumococcal antigen test for diagnosis of community-acquired pneumonia caused by *Streptococcus pneumoniae*. *J Clin Microbiol*. 2013;51:2303-10. [PMID: 23678060] doi:10.1128/JCM.00137-13
36. Alhethel AF, Albarrag AM, Somily AM, Eifan SA. H1N1 testing: a comparative study between the rapid BD(TM) Directigen EZ Flu A+B test and PCR. *J Pure Appl Microbiol*. 2014;8:2265-70.
37. Angoulvant F, Bellettre X, Houhou N, Dexpert JB, Morin L, Siriez JY, et al. Sensitivity and specificity of a rapid influenza diagnostic test in children and clinical utility during influenza A (H1N1) 2009 outbreak. *Emerg Med J*. 2011;28:924-6. [PMID: 20943835] doi:10.1136/emj.2010.098533
38. Avril E, Lacroix S, Vrignaud B, Moreau-Klein A, Coste-Burel M, Launay E, et al. Variability in the diagnostic performance of a bedside rapid diagnostic influenza test over four epidemic seasons in a pediatric emergency department. *Diagn Microbiol Infect Dis*. 2016;85:334-7. [PMID: 27139081] doi:10.1016/j.diagmicrobio.2016.03.015
39. Beckmann C, Hirsch HH. Diagnostic performance of near-patient testing for influenza. *J Clin Virol*. 2015;67:43-6. [PMID: 25959157] doi:10.1016/j.jcv.2015.03.024
40. Berthod D, Genton B, Hatz C, Blum J, de Vallière S. Ability of physicians to diagnose influenza and usefulness of a rapid influenza antigen test in febrile returning travelers: a randomized controlled trial. *Travel Med Infect Dis*. 2015;13:394-9. [PMID: 26358968] doi:10.1016/j.tmaid.2015.08.001
41. Binnicker MJ, Espy MJ, Irish CL, Vetter EA. Direct detection of influenza A and B viruses in less than 20 minutes using a commercially available rapid PCR assay. *J Clin Microbiol*. 2015;53:2353-4. [PMID: 25926484] doi:10.1128/JCM.00791-15
42. BinSaeed AA, Siddiqui AR, Mandil AM, Torchyian AA, Tayel SA, Shaikh SA, et al. The role of rapid testing and clinical decision in the diagnosis of human influenza A H1N1 infection. *Saudi Med J*. 2014;35:277-84. [PMID: 24623208]
43. Boyanton BL Jr, Almradi A, Mehta T, Robinson-Dunn B. Performance of the Directigen EZ Flu A+B rapid influenza diagnostic test to detect pandemic influenza A/H1N1 2009. *Diagn Microbiol Infect Dis*. 2014;78:360-2. [PMID: 24582319] doi:10.1016/j.diagmicrobio.2013.10.009
44. Bruening AH, van Dijk K, van Eijk HW, Koen G, van Woensel JB, Krusinga FH, et al. Evaluation of a rapid antigen detection point-of-care test for respiratory syncytial virus and influenza in a pediatric hospitalized population in the Netherlands. *Diagn Microbiol Infect Dis*. 2014;80:292-3. [PMID: 25241640] doi:10.1016/j.diagmicrobio.2014.08.010
45. Busson L, Mahadeb B, De Foor M, Vandenberg O, Hallin M. Contribution of a rapid influenza diagnostic test to manage hospitalized patients with suspected influenza. *Diagn Microbiol Infect Dis*. 2017;87:238-242. [PMID: 27939284] doi:10.1016/j.diagmicrobio.2016.11.015
46. Chiarella FC, Culebras E, Fuentes-Ferrer ME, Picazo JJ. Evaluation of the Alere i Influenza A&B assay for rapid identification of influenza A and influenza B viruses. *J Med Microbiol*. 2016;65:456-61. [PMID: 26967368] doi:10.1099/jmm.0.000249
47. Ciblak MA, Kanturvardar M, Asar S, Bozkaya E, Yenen OS, Badur S. Sensitivity of rapid influenza antigen tests in the diagnosis of pandemic (H1N1)2009 compared with the standard rRT-PCR technique during the 2009 pandemic in Turkey. *Scand J Infect Dis*. 2010;42:902-5. [PMID: 20662619] doi:10.3109/00365548.2010.502903
48. Cruz-Cañete M, Moreno-Pérez D, Jurado-Ortiz A, García-Martín FJ, López-Siles J, Olalla-Martín L. [Influenza virus in pediatrics. A reason for hospitalization]. *Enferm Infecc Microbiol Clin*. 2007;25:177-83. [PMID: 17335696]
49. DiMaio MA, Sahoo MK, Waggoner J, Pinsky BA. Comparison of Xpert Flu rapid nucleic acid testing with rapid antigen testing for the diagnosis of influenza A and B. *J Virol Methods*. 2012;186:137-40. [PMID: 22841669] doi:10.1016/j.jviromet.2012.07.023
50. Dunn J, Obuekwe J, Baun T, Rogers J, Patel T, Snow L. Prompt detection of influenza A and B viruses using the BD Veritor™ System Flu A+B, Quidel® Sofia® Influenza A+B FIA, and Alere BinaxNOW® Influenza A&B compared to real-time reverse transcription-polymerase chain reaction (RT-PCR). *Diagn Microbiol Infect Dis*. 2014;79:10-3. [PMID: 24582581] doi:10.1016/j.diagmicrobio.2014.01.018
51. Eggers M, Roth B, Schweiger B, Schmid M, Gregersen JP, Enders M. Comparison of the novel ResPlex III assay and existing techniques for the detection and subtyping of influenza virus during the influenza season 2006-2007. *Eur J Clin Microbiol Infect Dis*. 2012;31:1257-65. [PMID: 22012658] doi:10.1007/s10096-011-1437-1
52. Eggers M, Enders M, Terletskaia-Ladwig E. Evaluation of the Becton Dickinson rapid influenza diagnostic tests in outpatients in Germany during seven influenza seasons. *PLoS One*. 2015;10:e0127070. [PMID: 26011531] doi:10.1371/journal.pone.0127070
53. Gomez-Vinales C, Munguia R, Lara S, Ortiz G, Aldana C. [Outbreak of Influenza "A" H1N1 and the sensitivity and specificity of a rapid test]. *Enf Infecc Microbiol*. 2013;33:26-31.
54. González-Canudas J, Iglesias-Chiesa JM, Romero-Antonio Y, Chávez-Cortes C, Gay-Molina JG, Rivas-Ruiz R. [Cost-effectiveness in the detection of influenza H1N1: clinical data versus rapid tests]. *Rev Panam Salud Publica*. 2011;29:1-8. [PMID: 21390413]
55. Hassan F, Nguyen A, Formanek A, Bell JJ, Selvarangan R. Comparison of the BD Veritor System for Flu A+B with the Alere BinaxNOW influenza A&B card for detection of influenza A and B viruses in respiratory specimens from pediatric patients. *J Clin Microbiol*. 2014;52:906-10. [PMID: 24391198] doi:10.1128/JCM.02484-13
56. Hatano B, Goto M, Fukumoto H, Obara T, Maki T, Suzuki G, et al. Mobile and accurate detection system for infection by the 2009 pandemic influenza A (H1N1) virus with a pocket-warmer reverse-transcriptase loop-mediated isothermal amplification. *J Med Virol*. 2011;83:568-73. [PMID: 21328369] doi:10.1002/jmv.22031
57. Hazelton B, Gray T, Ho J, Ratnamohan VM, Dwyer DE, Kok J. Detection of influenza A and B with the Alere™ i Influenza A & B: a novel isothermal nucleic acid amplification assay. *Influenza Other Respir Viruses*. 2015;9:151-4. [PMID: 25728758] doi:10.1111/irv.12303
58. Hazelton B, Nedeljkovic G, Ratnamohan VM, Dwyer DE, Kok J. Evaluation of the Sofia Influenza A + B fluorescent immunoassay for the rapid diagnosis of influenza A and B. *J Med Virol*. 2015;87:35-8. [PMID: 24838873] doi:10.1002/jmv.23976

59. Hurtado JC, Mosquera MM, de Lazzari E, Martínez E, Torner N, Isanta R, et al. Evaluation of a new, rapid, simple test for the detection of influenza virus. *BMC Infect Dis.* 2015;15:44. [PMID: 25656393] doi:10.1186/s12879-015-0775-5
60. Jang JW, Ko SY, Byun MS, Sung HW, Lim CS. GENEDIA Multi Influenza Ag Rapid Test for detection and H1, H3, and H5 subtyping of influenza viruses. *J Clin Virol.* 2015;73:42-6. [PMID: 26540461] doi:10.1016/j.jcv.2015.10.014
61. Kawachi S, Matsushita T, Sato T, Nunoi H, Noguchi H, Ota S, et al. Multicenter prospective evaluation of a novel rapid immunochromatographic diagnostic kit specifically detecting influenza A H1N1 2009 virus. *J Clin Virol.* 2011;51:68-72. [PMID: 21324735] doi:10.1016/j.jcv.2011.01.007
62. Kenmoe S, Tchendjou P, Moyo Tetang S, Mossus T, Njankouo Ripa M, Guillet M, et al. Evaluating the performance of a rapid antigen test for the detection of influenza virus in clinical specimens from children in Cameroon. *Influenza Other Respir Viruses.* 2014;8:131-4. [PMID: 24266902] doi:10.1111/irv.12210
63. Koul PA, Mir H, Bhat MA, Khan UH, Khan MM, Chadha MS, et al. Performance of rapid influenza diagnostic tests (QuickVue) for influenza A and B infection in India. *Indian J Med Microbiol.* 2015;33 Suppl:26-31. [PMID: 25657152] doi:10.4103/0255-0857.148831
64. Leonardi GP, Wilson AM, Mitrache I, Zuretti AR. Comparison of the Sofia and Veritor direct antigen detection assay systems for identification of influenza viruses from patient nasopharyngeal specimens. *J Clin Microbiol.* 2015;53:1345-7. [PMID: 25609718] doi:10.1128/JCM.03441-14
65. Lewandowski K, Tamerius J, Menegus M, Olivo PD, Lollar R, Lee-Lewandowski E. Detection of influenza A and B viruses with the Sofia analyzer: a novel, rapid immunofluorescence-based in vitro diagnostic device. *Am J Clin Pathol.* 2013;139:684-9. [PMID: 23596120] doi:10.1309/AJCP7ZTLJCP3LLMA
66. Li X, Chen H, Wei J, Lv N, You L. The evaluation of colloidal gold immunochromatographic assay (GICA) for rapid diagnosis of influenza A disease. *Clin Chem Lab Med.* 2011;49:1533-7. [PMID: 21913792] doi:10.1515/CCLM.2011.235
67. Melchers WJG, Kuijpers J, Sickler JJ, Rahamat-Langendoen J. Lab-in-a-tube: Real-time molecular point-of-care diagnostics for influenza A and B using the cobas® Liat® system. *J Med Virol.* 2017;89:1382-1386. [PMID: 28213975] doi:10.1002/jmv.24796
68. Mese S, Akan H, Badur S, Uyanik A; Istanbul Rapid Test Study Group. Analytical performance of the BD Veritor™ system for rapid detection of influenza virus A and B in a primary healthcare setting. *BMC Infect Dis.* 2016;16:481. [PMID: 27612949] doi:10.1186/s12879-016-1811-9
69. Miarka M, Horban A, Maliszewska H, Bilinski P, Prus-Kowalczyk W. A clinical utility of a strip test for influenza A/B and comparison with detection by RT PCR. *Acta Biochim Pol.* 2014;61:485-7. [PMID: 25210936]
70. Mitamura K, Kawakami C, Shimizu H, Abe T, Konomi Y, Yasumi Y, et al. Evaluation of a new immunochromatographic assay for rapid identification of influenza A, B, and A(H1N1)2009 viruses. *J Infect Chemother.* 2013;19:633-8. [PMID: 23254398] doi:10.1007/s10156-012-0533-1
71. Moesker FM, van Kampen JJA, Aron G, Schutten M, van de Vijver DAMC, Koopmans MPG, et al. Diagnostic performance of influenza viruses and RSV rapid antigen detection tests in children in tertiary care. *J Clin Virol.* 2016;79:12-17. [PMID: 27045454] doi:10.1016/j.jcv.2016.03.022
72. Ndegwa LK, Emukule G, Uyeki TM, Mailu E, Chaves SS, Widdowson MA, et al. Evaluation of the point-of-care Becton Dickinson Veritor™ Rapid influenza diagnostic test in Kenya, 2013-2014. *BMC Infect Dis.* 2017;17:60. [PMID: 28077093] doi:10.1186/s12879-016-2131-9
73. Nguyen Van JC, Caméléna F, Dahoun M, Pilmis B, Mizrahi A, Lourtet J, et al. Prospective evaluation of the Alere i Influenza A&B nucleic acid amplification versus Xpert Flu/RSV. *Diagn Microbiol Infect Dis.* 2016;85:19-22. [PMID: 26899154] doi:10.1016/j.diagmicrobio.2015.11.012
74. Nitsch-Osuch A, Wozniak-Kosek A, Brydak LB. Accuracy of rapid influenza diagnostic test and immunofluorescence assay compared to real time RT-PCR in children with influenza A(H1N1)pdm09 infection. *Postepy Hig Med Dosw (Online).* 2012;66:752-7. [PMID: 23175329] doi:10.5604/17322693.1015040
75. Noh JY, Choi WS, Lee J, Kim HL, Song JY, Cheong HJ, et al. Clinical performance of the Sofia™ Influenza A+B FIA in adult patients with influenza-like illness. *Diagn Microbiol Infect Dis.* 2015;83:130-2. [PMID: 26184128] doi:10.1016/j.diagmicrobio.2015.05.016
76. Nolte FS, Gauld L, Barrett SB. Direct comparison of Alere i and cobas Liat influenza A and B tests for rapid detection of influenza virus infection. *J Clin Microbiol.* 2016;54:2763-2766. [PMID: 27582513]
77. Peci A, Winter AL, King EC, Blair J, Gubbay JB. Performance of rapid influenza diagnostic testing in outbreak settings. *J Clin Microbiol.* 2014;52:4309-17. [PMID: 25320225] doi:10.1128/JCM.02024-14
78. Peng Y, Wu J, Liu X, Wang J, Li W. Evaluation of Wondfo influenza A&B fast test based on immunochromatography assay for rapid diagnosis of influenza A H1N1. *Braz J Infect Dis.* 2013;17:247-50. [PMID: 23465599] doi:10.1016/j.bjid.2012.09.014
79. Rashid H, Shafi S, Booy R, El Bashir H, Ali K, Zambon M, et al. Influenza and respiratory syncytial virus infections in British Hajj pilgrims. *Emerg Health Threats J.* 2008;1:e2. [PMID: 22460211] doi:10.3134/ehjt.08.002
80. Rashid H, Shafi S, Haworth E, El Bashir H, Memish ZA, Sudhanva M, et al. Viral respiratory infections at the Hajj: comparison between UK and Saudi pilgrims. *Clin Microbiol Infect.* 2008;14:569-74. [PMID: 18373688] doi:10.1111/j.1469-0691.2008.01987.x
81. Rath B, Tief F, Obermeier P, Tuerk E, Karsch K, Muehlhans S, et al. Early detection of influenza A and B infection in infants and children using conventional and fluorescence-based rapid testing. *J Clin Virol.* 2012;55:329-33. [PMID: 22921515] doi:10.1016/j.jcv.2012.08.002
82. Reina J, Plasencia V, Leyes M, Nicolau A, Galmés A, Arbona G. [Comparison study of a real-time reverse transcription polymerase chain reaction assay with an enzyme immunoassay and shell vial culture for influenza A and B virus detection in adult patients]. *Enferm Infect Microbiol Clin.* 2010;28:95-8. [PMID: 19477042] doi:10.1016/j.eimc.2008.11.021
83. Reynders M, De Foor M, Maaroufi Y, Thomas I, Vergison A, Debulpaep S, et al. Prospective evaluation of Coris Inlu-A&B Respi-Strip and of BinaxNOW Influenza A&B assay against viral culture and real-time PCR assay for detection of 2009 pandemic influenza A/H1N1v in Belgian patients. *Acta Clin Belg.* 2012;67:94-8. [PMID: 22712164]
84. Ryu SW, Suh IB, Ryu SM, Shin KS, Kim HS, Kim J, et al. Comparison of three rapid influenza diagnostic tests with digital readout systems and one conventional rapid influenza diagnostic test. *J Clin Lab Anal.* 2017. [PMID: 28407318] doi:10.1002/jcla.22234
85. Sabetta J, Smardin J, Burns L, Barry K, Baisley C, Mahoney T, et al. Performance of rapid influenza diagnostic tests during two school outbreaks of 2009 pandemic influenza A (H1N1) virus infection—Connecticut, 2009. *MMWR Morb Mortal Wkly Rep.* 2009; 58:1029-32.
86. Sasaki T, Kubota-Koketsu R, Takei M, Hagihara T, Iwamoto S, Murao T, et al. Reliability of a newly-developed immunochromatography diagnostic kit for pandemic influenza A/H1N1pdm virus: implications for drug administration. *PLoS One.* 2012;7:e50670. [PMID: 23226350] doi:10.1371/journal.pone.0050670
87. Selim HS, Hashish MH. Performance of a rapid test versus real-time PCR for diagnosis of H1N1 swine flu. *J Egypt Public Health Assoc.* 2014;89:96-9. [PMID: 25162742] doi:10.1097/01.EPX.0000452224.96616.42
88. Selove W, Rao LV. Performance of rapid SOFIA Influenza A+B test compared to Luminex x-TAG respiratory viral panel assay in the diagnosis of influenza A, B, and subtype H3. *J Investig Med.* 2016; 64:905-7. [PMID: 26911275] doi:10.1136/jim-2016-000055
89. Shi W, Cui S, Peng X, Dwyer DE, Huang F, Wang Q, et al. Evaluation of the FluA-Ag rapid assay for detection of influenza A viruses

- of human, avian, and swine origin. *Clin Lab*. 2014;60:297-300. [PMID: 24660544]
90. Tenorio-Abreu A, Andaluz Ojeda D, Castro Hernandez B, Hernandez Porto M, Lecuona Fernandez M, Sierra Lopez A. Evaluation of a rapid antigen detection test for the diagnosis of emergency influenza A H1N1 pandemic in a pediatric population. *Acta Pediatr Esp*. 2012;70:251-3.
91. Tuttle R, Weick A, Schwarz WS, Chen X, Obermeier P, Seeber L, et al. Evaluation of novel second-generation RSV and influenza rapid tests at the point of care. *Diagn Microbiol Infect Dis*. 2015;81:171-6. [PMID: 25583129] doi:10.1016/j.diagmicrobio.2014.11.013
92. Watanabe M, Nukuzuma S, Ito M, Ihara T. Viral load and rapid diagnostic test in patients with pandemic H1N1 2009. *Pediatr Int*. 2011;53:1097-9. [PMID: 22004097] doi:10.1111/j.1442-200X.2011.03489.x
93. Yang JH, Huang PY, Shie SS, Huang CG, Tsao KC, Huang CT. Diagnostic capacity of rapid influenza antigen test: reappraisal with experience from the 2009 H1N1 pandemic. *J Microbiol Immunol Infect*. 2012;45:102-7. [PMID: 22177367] doi:10.1016/j.jmii.2011.09.027
94. Zazueta-García R, Canizalez-Roman A, Flores-Villaseñor H, Martínez-García J, Llausa-Vargas A, León-Sicairos N. Effectiveness of two rapid influenza tests in comparison to reverse transcription-PCR for influenza A diagnosis. *J Infect Dev Ctries*. 2014;8:331-8. [PMID: 24619265] doi:10.3855/jidc.3726
95. Agoritsas K, Mack K, Bonsu BK, Goodman D, Salamon D, Marcon MJ. Evaluation of the Quidel QuickVue test for detection of influenza A and B viruses in the pediatric emergency medicine setting by use of three specimen collection methods. *J Clin Microbiol*. 2006;44:2638-41. [PMID: 16825402]
96. Al Johani SM, Al Balawi M, Al Alwan B, Al Hefdhri R, Hajeer A. Validity of two rapid point of care influenza tests and direct fluorescence assay in comparison of real time PCR for swine of origin influenza virus. *J Infect Public Health*. 2011;4:7-11. [PMID: 21338954] doi:10.1016/j.jiph.2010.10.004
97. Alexander R, Hurt AC, Lamb D, Wong FY, Hampson AW, Barr IG. A comparison of a rapid test for influenza with laboratory-based diagnosis in a paediatric population. *Commun Dis Intell Q Rep*. 2005;29:272-6. [PMID: 16220863]
98. Bellmann-Weiler R, Beikircher B, Kurz K, Theurl I, Weiss G. Accuracy of bedside antigen tests in the diagnosis of new influenza A/H1N1v infection. *Clin Microbiol Infect*. 2011;17:235-7. [PMID: 20384708] doi:10.1111/j.1469-0691.2010.03235.x
99. Biggs C, Walsh P, Overmyer CL, Gonzalez D, Feola M, Mordechai E, et al. Performance of influenza rapid antigen testing in influenza in emergency department patients. *Emerg Med J*. 2010;27:5-7. [PMID: 20028996] doi:10.1136/emj.2009.078683
100. Boivin G, Hardy I, Kress A. Evaluation of a rapid optical immunoassay for influenza viruses (FLU OIA test) in comparison with cell culture and reverse transcription-PCR. *J Clin Microbiol*. 2001;39:730-2. [PMID: 11158137]
101. Boivin G, Côté S, Déry P, De Serres G, Bergeron MG. Multiplex real-time PCR assay for detection of influenza and human respiratory syncytial viruses. *J Clin Microbiol*. 2004;42:45-51. [PMID: 14715730]
102. Cheng XD, Yuan Q, Yue QH, Zheng QB, Ma YY, Yang BC, et al. Evaluation of a new rapid influenza A diagnostic test for detection of pandemic (H1N1) 2009 and seasonal influenza A virus. *J Clin Virol*. 2011;50:153-5. [PMID: 21051280] doi:10.1016/j.jcv.2010.10.002
103. Choi YJ, Kim HJ, Park JS, Oh MH, Nam HS, Kim YB, et al. Evaluation of new rapid antigen test for detection of pandemic influenza A/H1N1 2009 virus. *J Clin Microbiol*. 2010;48:2260-2. [PMID: 20357213] doi:10.1128/JCM.02392-09
104. Choi YJ, Nam HS, Park JS, Kim HJ, Park KB, Jeon MH, et al. Comparative analysis of the multiple test methods for the detection of Pandemic Influenza A/H1N1 2009 virus. *J Microbiol Biotechnol*. 2010;20:1450-6. [PMID: 21030832]
105. Crum-Cianflone NF, Blair PJ, Faix D, Arnold J, Echols S, Sherman SS, et al. Clinical and epidemiologic characteristics of an outbreak of novel H1N1 (swine origin) influenza A virus among United States military beneficiaries. *Clin Infect Dis*. 2009;49:1801-10. [PMID: 19911946] doi:10.1086/648508
106. Cruz AT, Demmler-Harrison GJ, Caviness AC, Buffone GJ, Revell PA. Performance of a rapid influenza test in children during the H1N1 2009 influenza A outbreak. *Pediatrics*. 2010;125:e645-50. [PMID: 20156902] doi:10.1542/peds.2009-3060
107. de la Tabla VO, Antequera P, Masiá M, Ros P, Martín C, Gazquez G, et al. Clinical evaluation of rapid point-of-care testing for detection of novel influenza A (H1N1) virus in a population-based study in Spain. *Clin Microbiol Infect*. 2010;16:1358-61. [PMID: 21382125] doi:10.1111/j.1469-0691.2010.03159.x
108. Faix DJ, Sherman SS, Waterman SH. Rapid-test sensitivity for novel swine-origin influenza A (H1N1) virus in humans [Letter]. *N Engl J Med*. 2009;361:728-9. [PMID: 19564634] doi:10.1056/NEJMc0904264
109. Foo H, Blyth CC, van Hal S, McPhie K, Ratnamohan M, Fennell M, et al. Laboratory test performance in young adults during influenza outbreaks at World Youth Day 2008. *J Clin Virol*. 2009;46:384-6. [PMID: 19828366] doi:10.1016/j.jcv.2009.09.019
110. Fuenzalida L, Blanco S, Prat C, Vivancos M, Dominguez MJ, Mödol JM, et al. Utility of the rapid antigen detection BinaxNOW Influenza A&B test for detection of novel influenza A (H1N1) virus. *Clin Microbiol Infect*. 2010;16:1574-6. [PMID: 20047602] doi:10.1111/j.1469-0691.2009.03160.x
111. Ganzenmueller T, Kluba J, Hilfrich B, Puppe W, Verhagen W, Heim A, et al. Comparison of the performance of direct fluorescent antibody staining, a point-of-care rapid antigen test and virus isolation with that of RT-PCR for the detection of novel 2009 influenza A (H1N1) virus in respiratory specimens. *J Med Microbiol*. 2010;59:713-7. [PMID: 20203216] doi:10.1099/jmm.0.017244-0
112. Gao F, Loring C, Laviolette M, Bolton D, Daly ER, Bean C. Detection of 2009 pandemic influenza A(H1N1) virus infection in different age groups by using rapid influenza diagnostic tests. *Influenza Other Respir Viruses*. 2012;6:e30-4. [PMID: 22114876] doi:10.1111/j.1750-2659.2011.00313.x
113. Ghebremedhin B, Engelmann I, König W, König B. Comparison of the performance of the rapid antigen detection actim Influenza A&B test and RT-PCR in different respiratory specimens. *J Med Microbiol*. 2009;58:365-70. [PMID: 19208888] doi:10.1099/jmm.0.004358-0
114. Gimeno C, Bravo D, Ocete D, Tormo N, Navalpotro D, Costa E, et al. Comparison of BinaxNOW Influenza A&B assay and real-time reverse transcription polymerase chain reaction for diagnosis of influenza A pandemic (H1N1) 2009 virus infection in adult patients. *Diagn Microbiol Infect Dis*. 2010;68:456-8. [PMID: 20884157] doi:10.1016/j.diagmicrobio.2010.07.010
115. Gooskens J, Swaan CM, Claas EC, Kroes AC. Rapid molecular detection of influenza outbreaks in nursing homes. *J Clin Virol*. 2008;41:7-12. [PMID: 18065263]
116. Gordon A, Videá E, Saborio S, López R, Kuan G, Reingold A, et al. Performance of an influenza rapid test in children in a primary healthcare setting in Nicaragua. *PLoS One*. 2009;4:e7907. [PMID: 19936063] doi:10.1371/journal.pone.0007907
117. Gröndahl B, Puppe W, Weigl J, Schmitt HJ. Comparison of the BD Directigen Flu A+B Kit and the Abbott TestPack RSV with a multiplex RT-PCR ELISA for rapid detection of influenza viruses and respiratory syncytial virus. *Clin Microbiol Infect*. 2005;11:848-50. [PMID: 16153263]
118. Hawkes M, Richardson SE, Ipp M, Schuh S, Adachi D, Tran D. Sensitivity of rapid influenza diagnostic testing for swine-origin 2009 a (H1N1) influenza virus in children. *Pediatrics*. 2010;125:e639-44. [PMID: 20156906] doi:10.1542/peds.2009-2669
119. Herrmann B, Larsson C, Zwegyberg BW. Simultaneous detection and typing of influenza viruses A and B by a nested reverse transcription-PCR: comparison to virus isolation and antigen detection by immunofluorescence and optical immunoassay (FLU OIA). *J Clin Microbiol*. 2001;39:134-8. [PMID: 11136761]
120. Karre T, Maguire HF, Butcher D, Graepler A, Weed D, Wilson ML. Comparison of Becton Dickinson Directigen EZ Flu A+B test against the CDC real-time PCR assay for detection of 2009 pandemic

- influenza A/H1N1 virus. *J Clin Microbiol*. 2010;48:343-4. [PMID: 19889893] doi:10.1128/JCM.02063-09
121. Kim YK, Uh Y, Chun JK, Kim C, Kim HY. Evaluation of new hemagglutinin-based rapid antigen test for influenza A pandemic (H1N1) 2009. *J Clin Virol*. 2010;49:69-72. [PMID: 20663709] doi:10.1016/j.jcv.2010.06.012
122. Kok J, Blyth CC, Foo H, Patterson J, Taylor J, McPhee K, et al. Comparison of a rapid antigen test with nucleic acid testing during cocirculation of pandemic influenza A/H1N1 2009 and seasonal influenza A/H3N2. *J Clin Microbiol*. 2010;48:290-1. [PMID: 19889892] doi:10.1128/JCM.01465-09
123. Landry ML, Cohen S, Ferguson D. Real-time PCR compared to Binax NOW and cytospin-immunofluorescence for detection of influenza in hospitalized patients. *J Clin Virol*. 2008;43:148-51. [PMID: 18639488] doi:10.1016/j.jcv.2008.06.006
124. Lee GC, Jeon ES, Kim WS, Le DT, Yoo JH, Chong CK. Evaluation of a rapid diagnostic test, NanoSign<sup>®</sup> Influenza A/B Antigen, for detection of the 2009 pandemic influenza A/H1N1 viruses. *Virology*. 2010;7:244. [PMID: 20849665] doi:10.1186/1743-422X-7-244
125. Lee HM, Park HK, Hwang HS, Chun MY, Pai HJ, Oh SM, et al. Diagnostic value of the rapid influenza antigen test for novel influenza A (H1N1). *Scand J Infect Dis*. 2011;43:43-6. [PMID: 20735325] doi:10.3109/00365548.2010.508463
126. Louie JK, Acosta M, Jamieson DJ, Honein MA; California Pandemic (H1N1) Working Group. Severe 2009 H1N1 influenza in pregnant and postpartum women in California. *N Engl J Med*. 2010;362:27-35. [PMID: 20032319] doi:10.1056/NEJMoa0910444
127. Lucas PM, Morgan OW, Gibbons TF, Guerrero AC, Maupin GM, Butler JL, et al. Diagnosis of 2009 pandemic influenza A (pH1N1) and seasonal influenza using rapid influenza antigen tests, San Antonio, Texas, April-June 2009. *Clin Infect Dis*. 2011;52 Suppl 1:S116-22. [PMID: 21342882] doi:10.1093/cid/ciq027
128. Ming C, Wei X, Biao A, Han H, Xuezheng L, Hong D, et al. Sensitivity assessment of rapid influenza diagnostic tests for the detection of the 2009 pandemic influenza A (H1N1) virus in clinical specimens. *Lab Med*. 2010;41:731-4.
129. Mizuike R, Sasaki T, Baba K, Iwamoto H, Shibai Y, Kosaka M, et al. Development of two types of rapid diagnostic test kits to detect the hemagglutinin or nucleoprotein of the swine-origin pandemic influenza A virus H1N1. *Clin Vaccine Immunol*. 2011;18:494-9. [PMID: 21228147] doi:10.1128/CVI.00269-10
130. Nilsson AC, Alemo B, Björkman P, Dillner L, Melhus A, Nilsson B, et al. Around-the-clock, rapid diagnosis of influenza by means of membrane chromatography antigen testing confirmed by polymerase chain reaction. *Infect Control Hosp Epidemiol*. 2008;29:177-9. [PMID: 18171307] doi:10.1086/526446
131. Noel G, Jachymczyk J, UTERS M, Laporte R, Jurquet AL, Parache C, et al. [Values of clinical signs and rapid diagnostic test in the diagnosis of influenza A (H1N1) new variant in pediatric emergency department]. *Arch Pediatr*. 2011;18:497-504. [PMID: 21489761] doi:10.1016/j.arcped.2011.02.017
132. Nougairede A, Ninove L, Zandotti C, de Lamballerie X, Gazin C, Drancourt M, et al. Point of care strategy for rapid diagnosis of novel A/H1N1 influenza virus. *PLoS One*. 2010;5:e9215. [PMID: 20174646] doi:10.1371/journal.pone.0009215
133. Nougairede A, Ninove L, Zandotti C, Thiberville SD, Gazin C, La Scola B, et al. Interim report on the A/H1N1 influenza virus pandemic in Marseille, France, April-November 2009. *Clin Microbiol Infect*. 2010;16:322-5. [PMID: 20121828] doi:10.1111/j.1469-0691.2010.03168.x
134. Poehling KA, Griffin MR, Dittus RS, Tang YW, Holland K, Li H, et al. Bedside diagnosis of influenza virus infections in hospitalized children. *Pediatrics*. 2002;110:83-8. [PMID: 12093950]
135. Poeppl W, Herkner H, Burgmann H, Pustelnik T, Mooseder G, Popow-Kraupp T, et al. Performance of the QuickVue Influenza A+B rapid test for pandemic H1N1 (2009) virus infection in adults. *PLoS One*. 2011;6:e28089. [PMID: 22145023] doi:10.1371/journal.pone.0028089
136. Pongthanapitth V, Sukasem C, Premchaiporn K, Srichantaratsamee C, Chantratita W. Clinical performance of three rapid diagnostic tests for influenza virus in nasopharyngeal specimens to detect novel swine-origin influenza viruses. *Infection*. 2011;39:105-11. [PMID: 21424855] doi:10.1007/s15010-011-0092-x
137. Rahman M, Vandermause MF, Kieke BA, Belongia EA. Performance of Binax NOW Flu A and B and direct fluorescent assay in comparison with a composite of viral culture or reverse transcription polymerase chain reaction for detection of influenza infection during the 2006 to 2007 season. *Diagn Microbiol Infect Dis*. 2008;62:162-6. [PMID: 18060723]
138. Rashid H, Shafi S, Haworth E, El Bashir H, Ali KA, Memish ZA, et al. Value of rapid testing for influenza among Hajj pilgrims. *Travel Med Infect Dis*. 2007;5:310-3. [PMID: 17870637]
139. Rouleau I, Charest H, Douville-Fradet M, Skowronski DM, De Serres G. Field performance of a rapid diagnostic test for influenza in an ambulatory setting. *J Clin Microbiol*. 2009;47:2699-703. [PMID: 19587306] doi:10.1128/JCM.00762-09
140. Ruest A, Michaud S, Deslandes S, Frost EH. Comparison of the Directigen flu A+B test, the QuickVue influenza test, and clinical case definition to viral culture and reverse transcription-PCR for rapid diagnosis of influenza virus infection. *J Clin Microbiol*. 2003;41:3487-93. [PMID: 12904343]
141. Sambol AR, Abdalhamid B, Lyden ER, Aden TA, Noel RK, Hinrichs SH. Use of rapid influenza diagnostic tests under field conditions as a screening tool during an outbreak of the 2009 novel influenza virus: practical considerations. *J Clin Virol*. 2010;47:229-33. [PMID: 20080438] doi:10.1016/j.jcv.2009.12.015
142. Sandora TJ, Smole SC, Lee GM, Chung S, Williams L, McAdam AJ. Test characteristics of commercial influenza assays for detecting pandemic influenza A (H1N1) in children. *Pediatr Infect Dis J*. 2010;29:261-2. [PMID: 19935118] doi:10.1097/INF.0b013e3181be9f9c
143. Stebbins S, Stark JH, Prasad R, Thompson WW, Mitruka K, Rinaldo C, et al. Sensitivity and specificity of rapid influenza testing of children in a community setting. *Influenza Other Respir Viruses*. 2011;5:104-9. [PMID: 21306573] doi:10.1111/j.1750-2659.2010.00171.x
144. Stein J, Louie J, Flanders S, Maselli J, Hacker JK, Drew WL, et al. Performance characteristics of clinical diagnosis, a clinical decision rule, and a rapid influenza test in the detection of influenza infection in a community sample of adults. *Ann Emerg Med*. 2005;46:412-9. [PMID: 16271670]
145. Suntarattiwong P, Jarman RG, Levy J, Baggett HC, Gibbons RV, Chotpitayasunondh T, et al. Clinical performance of a rapid influenza test and comparison of nasal versus throat swabs to detect 2009 pandemic influenza A (H1N1) infection in Thai children. *Pediatr Infect Dis J*. 2010;29:366-7. [PMID: 19949356] doi:10.1097/INF.0b013e3181c6f05c
146. Talbot HK, Williams JV, Zhu Y, Poehling KA, Griffin MR, Edwards KM. Failure of routine diagnostic methods to detect influenza in hospitalized older adults. *Infect Control Hosp Epidemiol*. 2010;31:683-8. [PMID: 20470035] doi:10.1086/653202
147. Uyeki TM, Prasad R, Vukotich C, Stebbins S, Rinaldo CR, Ferng YH, et al. Low sensitivity of rapid diagnostic test for influenza. *Clin Infect Dis*. 2009;48:e89-92. [PMID: 19323628] doi:10.1086/597828
148. Velasco JM, Montesa-Develos ML, Jarman RG, Lopez MN, Gibbons RV, Valderama MT, et al. Evaluation of QuickVue influenza A+B rapid test for detection of pandemic influenza A/H1N1 2009. *J Clin Virol*. 2010;48:120-2. [PMID: 20399140] doi:10.1016/j.jcv.2010.03.010
149. Zetti ZR, Wong KK, Haslina M, Ilina I. Preliminary evaluation of various rapid influenza diagnostic test methods for the detection of the novel influenza A (H1N1) in Universiti Kebangsaan Malaysia Medical Centre. *Med J Malaysia*. 2010;65:27-30. [PMID: 21265244]
150. Bruning A, Leeflang M, Vos J, Spijker R, de Jong MD, Wolthers KC, et al. Rapid tests for influenza, respiratory syncytial virus, and other respiratory viruses: a systematic review and meta-analysis. *Clin Infect Dis*. 2017. [PMID: 28520858] doi:10.1093/cid/cix461
151. Somerville LK, Ratnamohan VM, Dwyer DE, Kok J. Molecular diagnosis of respiratory viruses. *Pathology*. 2015;47:243-9. [PMID: 25764205] doi:10.1097/PAT.0000000000000240

152. Grimes DA, Schulz KF. Refining clinical diagnosis with likelihood ratios. *Lancet*. 2005;365:1500-5. [PMID: 15850636]
153. Ioannidis JP, Greenland S, Hlatky MA, Khoury MJ, Macleod MR, Moher D, et al. Increasing value and reducing waste in research design, conduct, and analysis. *Lancet*. 2014;383:166-75. [PMID: 24411645] doi:10.1016/S0140-6736(13)62227-8
154. Loeb M, Singh PK, Fox J, Russell ML, Pabbaraju K, Zarra D, et al. Longitudinal study of influenza molecular viral shedding in Hutterite communities. *J Infect Dis*. 2012;206:1078-84. [PMID: 22837493] doi:10.1093/infdis/jis450
155. Ng S, Lopez R, Kuan G, Gresh L, Balmaseda A, Harris E, et al. The timeline of influenza virus shedding in children and adults in a household transmission study of influenza in Managua, Nicaragua. *Pediatr Infect Dis J*. 2016;35:583-6. [PMID: 26910589] doi:10.1097/INF.0000000000001083
156. Caliendo AM, Gilbert DN, Ginocchio CC, Hanson KE, May L, Quinn TC, et al; Infectious Diseases Society of America (IDSA). Better tests, better care: improved diagnostics for infectious diseases. *Clin Infect Dis*. 2013;57 Suppl 3:S139-70. [PMID: 24200831] doi:10.1093/cid/cit578
157. Brendish NJ, Schiff HF, Clark TW. Point-of-care testing for respiratory viruses in adults: The current landscape and future potential. *J Infect*. 2015;71:501-10. [PMID: 26215335] doi:10.1016/j.jinf.2015.07.008



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